

## Inheritance of resistance in indica rice cultivar HUR 4-3 against bacterial leaf blight (*Xanthomonas oryzae* pv. *oryzae*)

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### Abstract

The mode of inheritance of resistance to *Xanthomonas oryzae* pv. *oryzae* strain BXO1 and BXO43 wild type of bacterial leaf blight disease of rice was studied in six generations of crosses of cultivars HUR 4-3 into PB-1460. The resistance cultivars PB-1460 showed 4.54% disease severity, while susceptible cultivar HUR 4-3 showed 53.01% disease severity against *Xanthomonas oryzae* pv. *oryzae*. The area under the disease progress curve (AUDPC) of resistance cultivar was 65.61 which are significantly less than the susceptible cultivar 649.90. The F<sub>1</sub> plants were observed to be resistant with an average disease severity 08.87% and AUDPC 110.26. The F<sub>2</sub> populations were classified in to four distinct classes on their genotypic ratio of 9:3:3:1 and phenotypically these populations were grouped in two distinct classes resistant and susceptible with their ratio of 13:3, respectively. However, B<sub>1</sub> and B<sub>2</sub> populations were classified in to two distinct classes as resistant (Resistant/ moderately resistant) and susceptible (moderately susceptible/ susceptible) in the ratio of genotypic 1:1:1:1 and 1:1 and phenotypically 1:1 and 1:0, respectively. The disease resistance occurs in the population is mainly due to cumulative effects of dominant and recessive two resistant genes i.e., *Xa21* and *xa13*. Chi-square analysis of the population was confirm the inheritance of resistance with their value are 1.24 and 0.66 indicating that the observed data are in line with expected ratio and follow Mendelian pattern of inheritance of resistance to bacterial leaf blight in B<sub>1</sub> and B<sub>2</sub> generations and modification in the Mendelian ratio of inheritance in the F<sub>2</sub> populations, it showed inhibitory gene action i.e., 13:3 that means dominant gene have cumulative effect of recessive gene.

### Highlights

- Two popular rice cultivars viz., HUR 4-3 and PB-1460 were used to study the inheritance of resistance for bacterial leaf blight disease.
- Disease resistance is governed by the synergistic effect of one dominant *Xa21* and one recessive gene *xa13*, and dominant gene has higher effect than the recessive gene.
- The resistance and susceptible reaction were observed in the progenies and it segregates in the ratio of 13 : 3 (Inhibitory gene action) in F<sub>2</sub>, 1 : 1 (test cross ratio) in B<sub>1</sub> and 1 : 0 in B<sub>2</sub> generation.

**Keywords:** Bacterial leaf blight, disease severity, inheritance, inhibitory gene action, AUDPC

Rice (*Oryza sativa* L.) is the most widely consumed stable food crop of Poaceae family for a large part of the world's human population, especially in Asia and over half of the global population depends on it for their feed (Sasaki, 2005 and Lal *et al.*, 2014). India, the second largest rice growing country has a production of 104.32 million tonnes and cultivation area of about 44.6 million hectares with an average productivity of 2.34 tonnes per hectare (Anonymous, 2013 and Rajasekar and Jeyakumar, 2014). It is, however, unfortunate that rice crop is threatened by considerable number of diseases (more than 40 diseases) of fungal, bacterial and viral origin, and that is one of the reasons for low yield of rice in the world including Asia, especially in India (Latif *et al.*, 2011, Barnwal *et al.*, 2013 and Singh *et al.*, 2013a). The diseases may appear at any growth stage of the plant, attacking the seed sown, root system, foliage, stalk, leaf sheath, inflorescence and even the developing grain (Virmani and Siddiq, 1998).

and to a great extent, the conduciveness of the environment in which it occurs (Gnanamanickam *et al.* 1999, Singh *et al.*, 2013a and Barnwal *et al.*, 2013).

Application of various chemicals to control the BLB is not an effective approach (Devadath, 1989). Therefore, exploitation of host plant resistance is considered the most effective, economical and environmentally safe measure for controlling BLB (Singh *et al.*, 2013a and b). The most effective approach to control BLB is using resistant varieties. This is due to the fact that the presence of different pathogenic races subsequently breaks the resistance of rice cultivars. The several attempts have been made to identify and characterise BLB resistance genes. Globally, more than thirty-eight genes (25 dominant and 13 recessive) conferring resistance against various strains of *X. oryzae* pv. *oryzae* have been identified (Chen *et al.* 2011) from diverse sources. Major resistance genes, including *Xa4*, *xa5*, *Xa7*, *xa13* and *Xa21* have been incorporated into rice cultivars, in order to

**Table 1. Parental description of *Indica* rice cultivar, its pedigree and features**

Name of cultivars	Parentage and Year of release	Specific features of <i>Indica</i> rice cultivars
HUR 4-3	Mutant of Lanjhi (2009)	Semi dwarf 90 - 100 cm, 135-140 days to maturity, grain type – long grain slender, fine resistant to leaf roller and brown plant hopper, susceptible to bacterial leaf blight disease; yield: 55-58 q/ ha
PB-1460 (Improve Pusa Basmati-1)	Pusa Basmati-1 x IRBB 60 (2008)	Semi dwarf 95-110 cm, 130-135 days to maturity, basmati rice grain type: long grain slender, very fine grain shape, elongation ratio 1.5: 1.8, strong aroma present, resistant to bacterial leaf blight, tolerant to sheath blight and blast disease due to presence of <i>Xa21</i> , <i>xa13</i> , <i>qSBR 11.1</i> and <i>Pi54</i> genes; yield: 45-48 q/ ha

Bacterial leaf blight (BLB) of rice caused by gram negative bacteria *Xanthomonas oryzae* pv. *oryzae*, is one of the most destructive diseases throughout the world, occurs mostly during the wet season when water overflows in rice fields. Bacterial leaf blight (BLB) can cause yield loss by 20 - 50% and as high as 80% and even 100% under very severe conditions (Agarwal *et al.* 2005 and Singh *et al.*, 2013b). Crop loss assessment studies have revealed that this disease reduces grain yield to varying levels, depending on the stage of the crop, degree of cultivar susceptibility

develop new resistant varieties (Perumalsamy *et al.*, 2010). However, the cultivars containing a single major resistance gene proved to be susceptible due to mutation in pathogen race. Recently, pyramiding of more than one major resistance gene has been proven to deliver durable resistance against BLB (Rajpurohit *et al.*, 2010). Therefore, this study was carried out to know the inheritance of bacterial leaf blight resistance in cultivar HUR 4-3, PB-1460 with their  $F_1$ 's and its segregating generation against *Xanthomonas oryzae* pv. *oryzae*.

**Table 2. Scale for bacterial leaf blight disease (Anonymous, 1996 and IRRI, 1996)**

Infection %	Score	Host response
0 %	0	Highly resistant (HR)
> 1-10 %	1	Resistant (R)
> 10-30 %	3	Moderately resistant (MR)
> 30-50 %	5	Moderately susceptible (MS)
> 50-75 %	7	Susceptible (S)
> 75-100 %	9	Highly susceptible (HS)

## Materials and Methods

### Experimental site

The experiment was carried out during two consecutive wet (*Kharif*) season and one dry (*Off-season*) season of 2012-2013 to 2013-2014. The two wet (*Kharif*) seasons trials were taken during June, 15<sup>th</sup> to November, 15<sup>th</sup> of 2012-2013 and 2013-2014 at the Agricultural Research Farm of Institute of Agricultural Sciences, Banaras Hindu University, Varanasi in Northern Gangetic Alluvial Plain of India (83°03'0"E longitude; 25°18'0"N latitude and an altitude of 128.93 m above sea level). The experimental soil was Gangetic alluvial (Ustochrept) with pH 7.6. It was moderately fertile-being low in organic carbon (0.39%); available nitrogen (198.4 kg ha<sup>-1</sup>); and medium in available phosphorus (15.7 kg ha<sup>-1</sup>) and potassium (215.4 kg ha<sup>-1</sup>). However, dry (*Off-season*) season trial was taken at Central Rice Research Institute, Cuttack, Odisha during December, 15<sup>th</sup> to May, 20<sup>th</sup> 2013 - 2014.

### Raising of rice seedlings and creating population

The experimental material of *Indica* rice (Varietal details describe in table-1) staggered sown at seven days interval in nursery bed during 15<sup>th</sup>, 22<sup>th</sup> and 29<sup>th</sup> June, 2012-2013. Twenty five days old single seedlings were transplanted in crossing block during seven days interval in July, 2012-2013, at Agricultural Research Farm, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, UP. The high yielding BLB susceptible cultivar HUR-4-3 (P<sub>1</sub>) taken as female crossed with bacterial leaf blight resistance cultivar PB-1460(P<sub>2</sub>) as male to produced five hundred seeds of F<sub>1</sub>'s. Only two hundred seeds of F<sub>1</sub>

and both parents were sown at seven days interval in nursery bed during 15<sup>th</sup>, 22<sup>th</sup> and 29<sup>th</sup> December, 2012-2013 at Research Farm of Central Rice Research Institute, Cuttack, Odisha. Forty days old single seedlings were transplanted in crossing block during seven days interval in January to February, 2013-2014. At flowering stage, fifty F<sub>1</sub> plants were selfed to produce approximately 1000 F<sub>2</sub> seeds and both the parents crossed with remaining F<sub>1</sub>'s (used as female) plants to generate 400-500 seeds of B<sub>1</sub> (F<sub>1</sub> with HUR 4-3 (P<sub>1</sub>)) and B<sub>2</sub> (F<sub>1</sub> with PB-1460 (P<sub>2</sub>)) backcross generation. Both recurrent and donor parents were again crossed within themselves to produce F<sub>1</sub> (HUR 4-3 x PB-1460) seeds. Six generations, namely, P<sub>1</sub> (susceptible parent), P<sub>2</sub> (resistance parent), F<sub>1</sub>, F<sub>2</sub>, B<sub>1</sub> and B<sub>2</sub> were raised in nursery bed during 15<sup>th</sup> June 2013-14 during *Kharif-Season*. Twenty one days old single seedling were transplanted in complete randomized block design under three replications in separate plots of 3 meter length spaced at 20 cm apart and the distance between plant to plant (15 cm) was maintained at Agricultural Research Farm, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi.

### Cultural Practices and Fertilizer Application

The experimental field was kept free from weeds by adopting manual weeding. The Fertilizers were applied @ 120 kg N, 60 kg P<sub>2</sub>O<sub>5</sub>, and 60 kg K<sub>2</sub>O per hectare under well irrigated condition. Half of N and full dose of P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O were used as basal in irrigated field after puddling. Remaining half of nitrogen was applied at a time of tillering as a top dressing. All the recommended cultural practices and plant protection measures (except bacterial leaf blight disease control) were followed for raising the healthy crop under irrigated conditions.

### Inoculum preparation and Inoculation on rice plants

The cultures of *Xanthomonas oryzae* pv. *oryzae* (strain BXO<sub>1</sub> and BX043 *wild type*) was obtained from Directorate of Rice Research, Hyderabad, India and sub cultured on peptone sucrose agar (PSA) medium (10 g l<sup>-1</sup> Sucrose, 10 g l<sup>-1</sup> Peptone, 1 g l<sup>-1</sup> glutamic

**Table 3. Comparison of per cent disease incidence (PDI) and Area Under Disease Progress Curve (AUDPC) on P<sub>1</sub>, P<sub>2</sub>, F<sub>1</sub>, F<sub>2</sub>, B<sub>1</sub> and B<sub>2</sub> progenies against *Xanthomonas oryzae* pv. *oryzae* strains BXO1 and BXO43 wild type**

Six generations with population size	Plant classified in Resistant: Susceptible group	Per cent Disease incidence (%) at 7 days interval of Inoculation				AUDPC value	Disease Score at 28 DAI	Disease Score at 28 DAI
		7 DAI ± SD	14 DAI ± SD	21 DAI ± SD	28 DAI ± SD			
P <sub>1</sub> (HUR 4-3) 300 plants	300 S	10.56 ± 1.16	22.04 ± 2.02	38.57 ± 1.88	53.91 ± 1.82	649.90	7	S
P <sub>2</sub> (PB-1460) 300 plants	300 R	1.54 ± 0.32	2.26 ± 0.33	4.07 ± 0.85	4.54 ± 0.62	65.61	1	R
F <sub>1</sub> s (200 plants)	196R : 4S	1.81 ± 0.37	4.67 ± 0.47	5.77 ± 0.50	8.81 ± 0.87	110.26	1	R
F <sub>2</sub> 's population 640 plants	113 R	1.62 ± 0.43	3.85 ± 0.70	4.90 ± 0.60	7.13 ± 0.53	91.89	1	R
	353 R/MR	6.08 ± 0.64	15.93 ± 1.18	16.51 ± 0.93	20.08 ± 1.23	318.68	3	MR
	55 MR	8.84 ± 1.30	19.43 ± 0.95	24.50 ± 2.18	25.10 ± 1.50	426.33	3	MR
	131 S	12.98 ± 2.13	22.07 ± 2.72	42.70 ± 1.48	55.98 ± 1.76	694.73	7	S
B <sub>1</sub> (F <sub>1</sub> x HUR 4-3) 300 plants	141 MR	7.75 ± 1.06	15.90 ± 0.66	20.42 ± 0.93	21.27 ± 1.12	355.76	3	R
	159 MS/S	10.86 ± 1.64	20.30 ± 1.77	33.17 ± 2.08	43.87 ± 1.16	565.80	5	MS
B <sub>2</sub> (F <sub>1</sub> x PB-1460) 300 plants	156 R	2.68 ± 1.02	4.03 ± 1.10	5.33 ± 1.15	6.89 ± 1.01	99.05	1	R
	144 MR	7.82 ± 1.18	16.93 ± 1.72	22.93 ± 1.37	27.60 ± 0.85	403.04	3	MR

SD: Standard deviation, DAI: Days after inoculation, R: Resistant, S: Susceptible, MR: Moderately resistant and MS: Moderately susceptible

acid, 0.5 g l<sup>-1</sup> K<sub>2</sub>HPO<sub>4</sub>, 0.25 g l<sup>-1</sup> MgSO<sub>4</sub>·7H<sub>2</sub>O, 16 g l<sup>-1</sup> bacto-agar and maintained it at pH 7.2 - 7.4 with 40% NaOH) for 2-3 days at 28 °C (Fahy and Persley, 1983). For pathogenicity test, leaf-clipping method was used for inoculation as described previously (Kauffman *et al.*, 1973) in the rice plants with *Xanthomonas oryzae* pv. *oryzae*. The test was conducted on fully developed leaves at the age of 55 days old rice plants after transplanting. The sterilised scissor dipped in bacterial suspension containing 10<sup>9</sup> cfu ml<sup>-1</sup>, was used for inoculation. Approximately 15-20 leaves of all plants were grasped in one hand and the top 1-3 inches of leaves were clipped off, simultaneously.

#### Disease Scoring, observation recorded and data collection

Following inoculation, the plants were observed and note it after every 24 hours time interval, the appearance of disease symptoms (lesion length was measured) and disease incidence were recorded

at 07, 14, 21 and 28 days after inoculation using a disease score index of 0 – 9 (IRRI, 1996). The disease scoring data were generated from disease score chart given in Table 2 in the Standard Evaluation System for rice (IRRI, 1996 and Anonymous, 1996) for disease appearance on 300 randomly selected plants from both parents, 200 plants from F<sub>1</sub> hybrids, 300 to 500 plants in segregating generations were used for hybrids and segregating generation to evaluate the response of host plant.

#### Statistical Analysis

The per cent disease incidence was calculated according to formula given by Gnanamanickam *et al.* (1999). The collected data for studied traits were pooled and standard statistical procedure (Singh and Chaudhary, 1995) and statistical software Windostat ver. 8.3 were applied for statistical analysis.  $\chi^2$  (chi-square) test for goodness-of-fit was used to study



Table 4. Inheritance of bacterial leaf blight resistance of six generation P<sub>1</sub>, P<sub>2</sub>, F<sub>1</sub>, F<sub>2</sub>, B<sub>1</sub> and B<sub>2</sub> progenies of Indica rice against pathogen *Xanthomonas oryzae* pv. *oryzae* strains BX01 and BX043 wild type

Six generations with population size	PDI at 28 DAI with SD (%)	Host response at 28 DAI	No. of plants observed	Gene combination (Xa21: Dominant in nature and xa21: recessive in nature)	Genotypic ratio	$\chi^2$ value	Phenotypic ratio (R : S)	$\chi^2$ value
P <sub>1</sub> (HUR 4-3) 300 plants	53.91 ± 1.82	S	300	xa21xa21Xa13Xa13	-	NS	-	NS
P <sub>2</sub> (PB-1460) 300 plants	4.54 ± 0.62	R	300	Xa21Xa21xa13xa13	-	NS	-	NS
F <sub>1,s</sub> (200 plants)	8.81 ± 0.87	MIR	194	Xa21xa21Xa13xa13	-	NS	-	NS
F <sub>2</sub> 's segregating population 640 plants	7.13 ± 0.53	R	113	Xa21Xa21xa13xa13:Xa21xa21xa13xa13	3	0.41		
	20.08 ± 1.23	MIR	347	Xa21Xa21Xa13Xa13:Xa21Xa21Xa13xa13 : Xa21xa21Xa13Xa13:Xa21xa21Xa13xa13	9	0.47	13 : 3 (509 : 131)	0.23
	25.10 ± 1.50	MIR	49	xa21xa21xa13xa13	1	2.03	R : S	
	55.98 ± 1.76	MS / S	131	xa21xa21Xa13Xa13:Xa13:xa21xa21Xa13xa13	3	1.01		1.01
	21.27 ± 1.12	R	143	Xa21xa21Xa13xa13:Xa21xa21Xa13Xa13	1	0.33	1 : 1 (143:157)	0.33
B <sub>1</sub> (F <sub>1</sub> x HUR 4-3) 300 plants	43.87 ± 1.16	MS / S	157	xa21xa21Xa13xa13:xa21xa21Xa13Xa13	1	0.33	R : S	0.33
	6.89 ± 1.01	R	158	Xa21Xa21xa13xa13:Xa21Xa21Xa13xa13	1:1	NS	1 : 0 All resistant	NS
B <sub>2</sub> (F <sub>1</sub> x PB-1460) 300 plants	27.60 ± 0.85	MIR	142	Xa21xa21xa13xa13:Xa21xa21Xa13xa13	1:1	NS		

PDI: Per cent disease incidence, SD: Standard deviation, DAI: Days after inoculation and gene (bold letters) showed resistance



Figure 1(a). HUR 4-3 a portion of normal field view, Bacterial leaf blight symptoms appeared in the field indicated by arrow.



(b) Comparison of lesion size pattern between leaves carrying either single gene, double gene or no gene of resistance



Figure 1 (c). Field plot inoculated with BLB pathogen (and arrow indicate appearance of lesion on the leaves) in cultivars HUR 4-3; (d) Panicle (F2 population) showing typical symptom of BLB after 21 days of inoculation. (e) Healthy plants of PB 1460 (un-inoculated) showing resistance against BLB under field condition. (f) B2 (H UR 4-3 x PB1460/ + PB1460) plant showing resistance against BLB after 21 days of inoculation

the Mendelian pattern of inheritance of resistance in bacterial leaf blight.

### Results and Discussion

The parental lines with their segregating population were evaluated under field condition for their resistance to bacterial leaf blight (BLB) using artificial inoculation of two different isolates of *Xoo*. One of these isolates, called *BXO1*, belongs to pathotype Ib. It is widely distributed *Xoo* pathotype of *Xanthomonas species* in all over India (Yoshitola *et al.*, 1997). The *Indica* cultivars PB-1460 was resistance due to presence of resistance genes *Xa21* and *xa13* which showed 4.54 % disease severity, while HUR

4-3 susceptible due to absence of these genes and showed 53.91 % disease severity against *Xanthomonas oryzae* pv. *oryzae* under epiphytotic condition (Table 3). The initial symptoms of bacterial leaf blight *viz.* linear yellow to straw coloured stripes with wavy margins, generally on both edges of leaf, rarely on one edge was observed with variable intensities in cultivar HUR 4-3 (Figure 1a). These finding were good agreement with the earlier report by Kihupi *et al.* (2001) and Singh *et al.* (2013b). The dominant gene *Xa21* for bacterial leaf blight resistance was introgressed in the cultivar PB-1460 from the isogenic line IRBB 60, and in the isogenic line IRBB 60, *Xa21* gene was introgressed from the wild source *Oryza*

*longistiminata* (Kihupi *et al.*, 2001 and Sunderam *et al.*, 2009).

The area under the disease progress curve of resistance cultivar (PB-1460) was 65.61 which are significantly less than the susceptible cultivar (HUR 4-3) 649.90. The  $F_1$  plants were observed to be resistant to moderately resistant when screened with a virulent isolate of *Xanthomonas oryzae* pv. *oryzae* strains, BXO1 and BXO43 wild type with average per cent disease severity 08.81% and AUDPC 110.26. This indicates the possible involvement of dominance gene(s) in governing the resistance. The  $F_1$  plants of cross HUR 4-3 x PB-1460 were selfed to produce  $F_2$  mapping population, these  $F_2$  mapping population individually scored and could be classified in to four distinct classes on the basis of their genotypic ratio of 9:3:3:1 with their gene combination (9  $Xa21Xa21Xa13Xa13/ Xa21Xa21Xa13xa13/ Xa21xa21Xa13Xa13/ Xa21xa21Xa13xa13$  : 3  $Xa21Xa21xa13xa13/ Xa21xa21xa13xa13$  : 3  $xa21xa21Xa13Xa13/ xa21xa21Xa13xa13$  : 1  $xa21xa21xa13xa13$ ) and two distinct classes on the basis of their phenotypically performance as resistant and susceptible with their ratio of 13:3, respectively (Table-4 and Figure 1b). Out of 640  $F_2$  plants, 503 plants were resistant and 131 plants showed susceptible reaction against BLB in the ratio of 13 : 3 with  $\chi^2 = 1.24$ ,  $P > 0.05$  indicating that observed data are in accordance with expected ratio. These results showed the *xa13* + *Xa21* gene combination was preferred in parental and their segregating generations to exploit the synergistic effects of this combination in preventing BLB infection. Similar findings were also reported by Sidhu *et al.* (1978), Kihupi *et al.* (2001) and Natarajkumar *et al.* (2008).

In backcross population, three hundred plants of each backcross were screened against isolates of *Xanthomonas oryzae* pv. *oryzae*. In first backcross population  $B_1$  ( $F_1$  x HUR 4-3) and in second backcross  $B_2$  ( $F_1$  x PB-1460) populations were individually disease scored and grouped in two distinct classes (Figure 1d and e). The per cent disease severity (PDI) and AUDPC value of the both backcross populations were 21.27% to 43.87% and 6.89% to 27.60% and 355.76

to 565.80 and 99.05 to 403.04, respectively (table-3). These population were again classified as resistant / moderately resistant and moderately susceptible / susceptible group on the basis of their genotypic ratio and gene combination 1:1 (1  $Xa21xa21Xa13xa13/ Xa21xa21Xa13Xa13$ : 1  $xa21xa21Xa13xa13/ xa21xa21Xa13Xa13$ ) and 1:1 (1  $Xa21Xa21xa13xa13/ Xa21Xa21Xa13xa13$ : 1  $Xa21xa21xa13xa13/ Xa21xa21Xa13xa13$ ), respectively. However, on the basis of phenotypic performance of both backcrosses they again classified in to diverse grouped *i.e.*, resistant and moderately susceptible/ Susceptible.  $B_1$  revealed 1 (resistant) : 1 (susceptible) ratio, thereby indicating that the 143 plants have single dominant resistance gene, While,  $B_2$  population were exhibited 1 (resistant) : 0 (susceptible) ratio, it indicates the resistance is governs by single dominant and recessive two resistance genes or sometimes it governs by only single dominant resistance gene (Figure 1f). These finding were in agreement with earlier report by Sundaram *et al.* (2009). Out of 300 plants of each backcross population, 143 resistant and 157 susceptible were found in  $B_1$  generation and all are resistant were found in  $B_2$  generation  $\chi^2 = 0.66$ ,  $P > 0.05$  and  $\chi^2 =$  Non Segregating (NS),  $P > 0.05$ , respectively indicating that observed data are in agreement with the expected ratio in backcross generation.

The  $F_2$ ,  $B_1$  and  $B_2$  progenies were segregates in the ratio 13 : 3, 1 : 1 and 1 : 0 which is mainly due to cumulative or synergistic effects of both resistance genes *i.e.*, one dominant gene *Xa21* and one recessive *xa13* resistant genes. Interestingly, when we compared the level of bacterial leaf blight resistance of cultivar HUR 4-3 with PB-1460, it was observed that the latter showed a higher degree of resistance as compared to PB - 1460. This could be due to the effect of different genetic backgrounds of HUR 4-3 and PB-1460, which may be responsible for modulating the level of resistance.

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## References

- Agarwal, P.K., Sidu, G.S., Gosal, S.S. 2005. Induction of bacterial blight resistance in elite Indian rice (*Oryza sativa* L.) cultivars using gamma irradiation and ethyl methane sulfonate. *Mutation Breeding Newsletter and Reviews* **1**: 17-18.
- Anonymous, 1996. Ministry of Food, Agriculture and Livestock. Food, Agriculture and livestock Division (Economic Wing) Islamabad. *Agriculture Statistics of Pakistan* pp. 13-17.
- Anonymous, 2011. *Directorate of Economics and Statistics*. New Delhi, Govt. of India.
- Barnwal, M.K., Kotasthane, A., Magculia, N., Mukherjee, P.K., Savary, S., Sharma, A.K., Singh, H.B., Singh, U.S., Sparks, A.H., Variar, M., Zaidi, N. 2013. A review on crop losses, epidemiology and disease management of rice brown spot to identify research priorities and knowledge gaps. *European Journal of Plant Pathology* **136**:443-457.
- Chen, S., Liu, X., Zeng, L., Ouyang, D, Yang, J., Zhu, X. 2011. Genetic analysis and molecular mapping of a novel recessive gene *xa34(t)* for resistance against *Xanthomonas oryzae* pv. *oryzae*. *Theoretical Applied Genetics* **122**: 1331-1338.
- Devadath, S. 1989. Chemical control of bacterial leaf blight of rice. In: IRRI (ed.) *Bacterial Blight of Rice* IRRI, Manila, Philippines, pp 89-98.
- Fahy, P.C., Persley, G.J. 1983. *Plant bacterial diseases: a diagnostic guide* Academic Press, New York, p. 393.
- Gnanamanickam, S.S., Priyadarisini, V.B., Narayanan, N.N., Vasudevan, P., Kavita, S. 1999. An overview of bacterial blight disease of rice and strategies for its management. *Current Science* **77**(11): 1435-1444.
- IRRI, 1996. *Standard Evaluation System for Rice* 4th Edn., International Rice Research Institute, Manila, Philippines.
- Kauffman, H.E., Reddy, A.P.K., Heish, S.P.Y., Marca, S.D. 1973. An improved technique for evaluating resistance of rice varieties to *Xanthomonas oryzae*. *Plant Disease Reports* **57**: 537-541.
- Kihupi, A.N., Angeles, E.R., Khush, G. S. 2001. Genetic analysis of resistance to bacterial blight, *Xanthomonas oryzae* pv. *oryzae*, in rice, *Oryza sativa* L. *Euphytica* **117**: 39-46
- Lal, D., Shashidhar, H.E., Ramanjini, P.H.G., Ashok, T.H. 2014. Callus Induction and Regeneration from *In vitro* anther culture of rice (*Oryza sativa* L.). *International Journal of Agriculture, Environment and Biotechnology* **7**(2): 213-218.
- Latif, M.A., Badsha, M.A., Tajul, M.I., Kabir, M.S., Rafii, M.Y., Mia, M.A.T. 2011. Identification of genotypes resistant to blast, bacterial leaf blight, sheath blight and tungro and efficacy of seed treating fungicides against blast disease of rice. *Scientific Research and Essays* **6**(13): 2804-2811.
- Natarajkumar, P., Sujatha, K., Laha, G.S., Mishra, B., Viraktamath, B.C., Srinivasarao, K., Hari, Y., Balachandran, S.M., Sundaram, R.M. 2008. Inheritance of bacterial blight resistance in the landrace Acc. 32753. *Rice Genetics Newsletter* **25**: 62-63.
- Perumalsamy, S., Bharani, M., Sudha, M., Nagarajan, P., Arul, L., Saraswathi, R., Balasubramanian, P., Ramalingam, J. 2010. Functional marker-assisted selection for bacterial leaf blight resistance genes in rice (*Oryza sativa* L.). *Plant Breeding* **129**: 400-406.
- Rajasekar, N., Jeyakumar, P. 2014. Differential response of trifloxystrobin in combination with tebuconazole on growth, nutrient uptake and yield of rice (*Oryza sativa* L.). *International Journal of Agriculture, Environment and Biotechnology* **6**(1): 87-93.
- Rajpurohit, D., Kumar, R., Kumar, M., Paul, P., Awasthi, A., Basha, P.O., Puri, A., Jhang, T., Singh, K., Dhaliwal, H.S. 2010. Pyramiding of two bacterial blight resistance and a semi dwarfing gene in Type 3 Basmati using marker-assisted selection. *Euphytica* **178**: 111-126.
- Sasaki, T. 2005. The map-based sequence of the rice genome. *Nature* **436**: 793-800.
- Sidhu, G.S., Khush, G.S., Mew, T.W. 1978. Genetic analysis of bacterial blight resistance in seventy-four cultivars of rice. *Theoretical Applied Genetics* **53**: 105-111.
- Singh, A.K., Sarma, B.K., Singh, P.K., Nandan, R. 2013b. Screening of rice (*Oryza sativa* L.) germplasms against *Xanthomonas oryzae* pv. *oryzae*. *Journal of Eco-friendly Agriculture* **8**(1): 86-88.
- Singh, M.K., Singh, P., Singh, R.P., Mohapatra, C. 2013a. Association analysis for yield and quality attributes in Indica rice and screening of hybrids against blast disease (*Magnaporthe grise* Barr.). *Journal of Plant Science*.
- Singh, R.K., Chaudhary, B.D. 1995. *Biometrical Methods in Quantitative Genetic analysis*. Kalyani Publisher, Ludhiana, New Delhi, ISBN: 8176633070, p. 342.
- Sundaram, R.M., Vishnupriya, M.R., Laha, G.S., Rani, N.S., Rao, P.S., Balachandran, S.M., Reddy, G.A., Sarma N.P., Sonti, R.V. 2009. Introduction of bacterial blight resistance into Triguna, a high yielding, mid-early duration rice variety. *Biotechnology Journal* **4**(3): 400-407.
- Virmany, S.S., Siddiq, E.A. 1998. Advances in hybrid rice technology. Proc. 3rd Intl on hybrid rice. IRRI, The Philippines.