

Arbuscular Mycorrhizal Fungi: A Potential Tool for Enhancing Crop Productivity in Salt Affected Soil

Sudeshna Bhattacharjya^{1*}, Debarati Bhaduri² and Asha Sahu¹

¹ICAR-Indian Institute of Soil Science, Bhopal-462038, Madhya Pradesh, India

²ICAR-National Rice Research Institute, Cuttack-753006, Odisha, India

*Corresponding author: sudeshna.bb@outlook.com (ORCID ID: 0000-0003-2883-8043)

Paper No. 753

Received: 21-07-2018

Accepted: 26-11-2018

ABSTRACT

AMF have been mentioned in several literatures for long time for their beneficial role in crop growth and productivity. However, recently they are talked about for efficient abiotic stress management which received equal attention among researchers. Salinity is such an abiotic stress faced by crop plants that can be minimized by assistance of AMF. In this article, we have discussed the possible mechanism and the identified species of AMF for abatement of salinity stress of plants by citing some suitable examples, apart from the harmful effects in crop plants' functioning due to salinity usually take place. Though there is lots of scope of further research, this illustrative piece of information may generate interest among farmers and other common people.

Highlights

- AM fungi are potential bio-remediation tool to alleviate salt stress in crops grown in saline soil for sustaining crop productivity in an eco-friendly way.

Keywords: Arbuscular Mycorrhizal Fungi (AMF), Salt stress, Salinity, Crop growth

Saline soil is characterized by higher concentration of soluble salts of Ca^{2+} , Mg^{2+} , K^+ , Na^+ , Cl^- , NO_3^- , SO_4^{2-} and CO_3^{3-} etc. Sodic soil is dominated by Na^+ salt and saline-sodic soil that have both high salt of Ca^{2+} , Mg^{2+} , K^+ as well as Na^+ . About 7% of the total land area of world (952.2 million ha) is salt affected. Salt affected which also constitutes nearly 33% of the potential arable land. In India, the salt affected soils account for 6.727 million ha (2.1%) of geographical area of the country. Furthermore, patches of salt affected soils are also emerging due to intensive crop cultivation and excessive exploitation of ground water.

Soil health is one of the key indicators of ecosystem functioning and the ecosystem more resilient to changes. The high salt concentrations adversely affect different soil processes and/or soil properties and in turn environmental conditions. Soil salinity pose a major threat to agricultural productivity as it

affects the establishment, growth and development of plants leading to significant loss in productivity (Giri *et al.* 2003; Mathur *et al.* 2007). To alleviate salt stress different techniques *viz.* introduction of transgenic salt tolerant crop variety, leaching, tile drainage etc. have already been employed. Although successful, large scale application of these technique is highly-priced and beyond the reach of developing nations (Evelin *et al.* 2009). Therefore, there is a constant need to generate some eco-friendly as well as effective measure to address this issue.

Plants, in their natural environment are colonized both by external and internal microorganisms. Arbuscular mycorrhizal fungi (AMF) are broad-spectrum and non-specific microorganism that are known to colonize 85% of terrestrial plant species including halophytes, hydrophytes and xerophytes (Pal *et al.* 2013). Several studies have reported the



ability of plants to be colonized by AMF in saline conditions, salt marshes and at neutral or moderate alkaline soil pH (Garcia and Mendoza, 2007). The soil borne or extra-matrical hyphae of AMF increase effective absorptive surface area of the plant roots resulting in higher nutrient uptake by crop in nutrient-poor soil (Pal *et al.* 2017). Besides improving nutritional status; they are also capable of enhancing physiological processes like water absorption capacity of plants by augmenting root hydraulic conductivity and favorably adjusting the osmotic balance and composition of carbohydrates (Rosendahl and Rosendahl, 1991; Feng *et al.* 2002). Thus, they alleviate the harmful effects of excess salt accumulated in the root (Dixon *et al.* 1993). Therefore, AMF found in environments with high salt concentrations could be a potential tool for bioremediation of salt affected soils and for augmenting crop productivity in an environmentally safe way.

Salinity effects on crops: an overview

High soil salinity impacts the growth of numerous plant species especially glycophytes (salt-sensitive compared to salt-tolerant halophytes species). Most of the agricultural crops fall in the category of glycophyte which exhibit a spectrum of responses under salt stress. Salinity not only decreases the agricultural production of most crops, but also, affects soil physico-chemical properties, and ecological balance of the area leading to low economic returns and soil erosions. The specific effects comprises of the following:

- ♦ Excessive accumulation of sodium in cell walls can results into osmotic stress and cell death (Munns, 2002).
- ♦ High salt concentration in soil can disturb the nutrient balance in the plant or interfere with the uptake of some nutrients (Blaylock *et al.* 1994; Chakraborty *et al.* 2015).
- ♦ Substantial decrease in leaf area, chlorophyll content and stomatal conductance, leading to less efficient photosystem (PS-II) (Netondo *et al.* 2004).
- ♦ Significant reduction in phosphorus (P) uptake by plant due to precipitation of phosphate ions as Ca-phosphate (Bano and Fatima 2009; Bhaduri *et al.* 2016).

- ♦ Salinity also impedes reproductive development by impairing micro-sporogenesis and stamen filament elongation, ovule abortion and senescence of fertilized embryos.
- ♦ Retarded supply of photosynthetic assimilates or hormones to the growing tissues (Ashraf, 2004).
- ♦ Soil salinity also results in ion toxicity where ratio of K^+ to Na^+ changes leading to replacement of K^+ by Na^+ in biochemical reactions. Because of this several enzymatic activities as well as binding tRNA to ribosomes consequently protein synthesis where K^+ is indispensable are inhibited. Moreover high level of Na^+ and Cl^- induced conformational changes in proteins. Recent reports also show that salinity adversely affects plant growth and development by hindering enzyme activity (Seckin *et al.* 2009), DNA, RNA, protein synthesis and mitosis (Tabur and Demir, 2010; Javid *et al.* 2011)
- ♦ Ion toxicity and osmotic stress could also cause metabolic imbalance, which in turn leads to oxidative stress (Chinnusamy *et al.* 2006; Chakraborty *et al.* 2016).
- ♦ Activities of cyclin and cyclin-dependent kinases are reported to be hindered by soil salinity resulting in fewer cells in the meristem, and limited growth. The activity of cyclin-dependent kinase is diminished also by post-translational inhibition during salt stress.

Role of AMF towards crop growth and nutrition

Interaction between plants and the AMF has a special relevance from ecological point of view, as this association brings about array of physiological changes *viz.* changes in concentration of growth regulating substances, increased leaf area and enhanced photosynthetic rate, photosynthetic partitioning to shoot and roots, improved nutritional status of host tissues facilitating the sustenance, growth and development of plant in the nature. There are number of prioritized issues in sustainable crop production like plant growth and nutrition, soil structuring, stress tolerance and survival, and increasing nutrient use efficiency etc. that have often been achieved by successful intervention of AMF (Yadav *et al.* 2017). The major effects of AM fungi symbiosis on crop growth are broadly as follows:

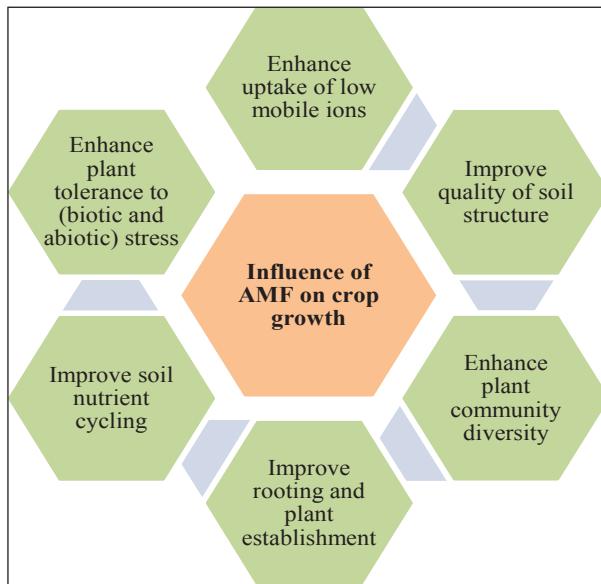


Fig. 1: The major effects of AMF symbiosis on crop growth

Plants, in their natural environment are colonized a wide array of both external and internal microorganisms. Arbuscular mycorrhizal fungi (AMF) are omnipresent and very common among the soil microorganisms dwelling in the rhizosphere and colonizes wide group of plant species in nature. AMF associations with plants are comprised of three main structures; viz: hyphae that performs as extended root surface scavenging the nutrients from the zone beyond the reach of plant roots (Hodge 2000); the second one is vesicles within the root, acting as storage organs, especially for lipids (Hirsch and Kapulnik 1998) and the third one is arbuscules which are branched intercellular structures and works as interface for phosphorus and other nutrient exchange on the root system (Smith et al. 2000). These fungi, being an important as well as integral component of the natural ecosystem and are found to exist in stressed environments such as saline soils (Giri et al. 2003). Although AMF are not uncommon in saline soils, their growth and colonization in plant roots has a fair chance to be affected by the excess of salt concentrations, which generally restrict the microbial growth and activity (Juniper and Abbott, 2006). Over the years, AM symbiosis has not only demonstrated the enhanced plant growth and biomass, but also imparted partial resistance to salinity and in a range of host plants such as maize, clover, tomato, cucumber, and lettuce (Rosendahl and Rosendahl, 1991; Ruiz-Lozano and Azcón, 1996; Al-Karaki et al. 2001; Feng et al. 2002).

A number of studies from last few decades have been carried out for investigating the role of AMF towards the enhanced crop growth and better nutrition. These kinds of studies varied from field crops to horticultural crops to forest sp. The results mostly indicated that symbiosis between AMF and plant root emerged to be beneficial in terms of expressing the physiological behaviour (photosynthetic efficiency, water use efficiency) of plants and nutrient uptake and accumulation in a better way (Porcel et al. 2012) that often reflect in increased plant biomass of the host plants. Altogether it was found that AMF-associated plants sustained their growth and yield, and this effect was more pronounced in salt-stress condition. Visually better response in plant biomass differentiated the positive role of AMF over non-inoculated plants (Sannazzaro et al. 2007; Zuccarini and Okurowska, 2008; Hajiboland et al. 2010). Moreover, better biomass and physiological activities of wheat with root colonization by *Glomus claroideum* (Beltrano and Ronco 2008) and strawberry plants by *Glomus* sp. (Borkowska 2002) were observed under drought stress as well. Antagonistic relationship between plant parasitic nematode and AMF also showed beneficial effect towards the crop growth by enhancing nutrient uptake (Calvet et al. 1995). Hence it can be expected that AMF substantially ameliorate both biotic and abiotic stresses, deploying an adequate vigour of crop plants.

Uptake of essential nutrients like P, K and micronutrients like Fe, Zn, Cu is often researched and highlighted under influence of AMF both under saline and non-saline soil conditions. However, when a plant is already facing a stress and the nutritional balance is already disturbed, the favorable effect created by AMF symbiosis is more pertinent in this regard. It is speculated that higher K^+ accumulation by tissues of mycorrhizal plants under salt stress may help to maintain a high K/Na ratio, which further may help into normalization of various enzymatic processes and protein synthesis (Porcel et al. 2012). Similar results showed in *Acacia nilotica* plants when colonized by *Glomus fasciculatum* was found to have a higher K^+ ion concentration in both root and shoot tissues even at all salinity levels (Giri et al. 2007). Improvement of P-ion uptake and accumulation in plants by better absorption of hyphal network articulated by AMF



had repeatedly been evidenced over the decades (Plenchette and Duponnis, 2005; Sharifi *et al.* 2007).

Ameliorative mechanisms involved in abatement of salt stress in crops

Salt stress can potentially cause a severe setback to plant growth, yield, physiological attributes and nutrient uptake behaviour. However, there are number of studies carried out to assess the role of AMF for alleviating the salt stress in crop plants, and most of them reported a positive outcome. The following figure (Fig. 2) depicts in brief the mechanisms imparted by AMF for abatement of salt stress to crops.

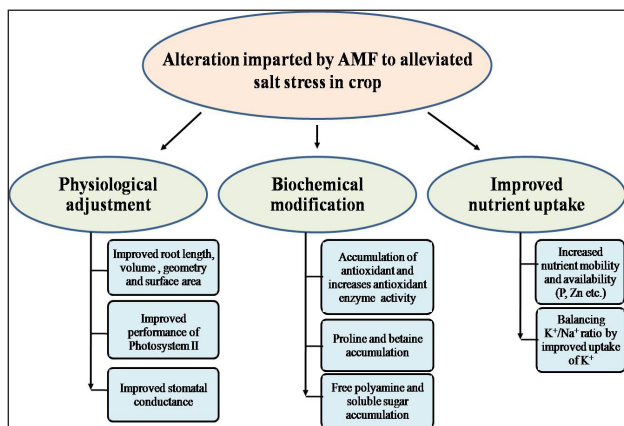


Fig. 2: The ameliorative mechanisms involved in alleviating salt stress in crops

Morpho-physiological changes

Salt stress can potentially cause a severe setback to plant growth, yield, physiological attributes and nutrient uptake behaviour. However, there are number of studies carried out to assess the role of AMF for alleviating the salt stress in crop plants, and most of them reported a positive outcome. Morphological and physiological characteristics of plants are keys to address any abiotic (salt) stress management hence most integral part of such experiments.

Many examples can be found in horticultural crops regarding useful inoculation of AMF. Association of AMF showed beneficial effect on root morphology of *Citrus tangerine* seedlings and enhanced the characters like root length, root projected area, root surface area and root volume under salinity (Wu *et al.* 2010a). Mycorrhizal colonization also caused improvement in fruit fresh weight, fruit number

and fruit yield of salt-stressed tomato plants (Al-Karaki, 2000; Latef and Chaoxing, 2011). Improved growth, yield and quality of fruits of *Cucurbita pepo* plants was also noticed when colonized by *Glomus intraradices* in salinity stress (Colla *et al.* 2008). Olive plants inoculated with *Glomus mosseae* helped to survive the plants better in salt stressed condition in terms of enhanced root and shoot growth, and lesser biomass reduction (Porrás-Soriano *et al.* 2009). Among field crops, AMF inoculated maize plants showed to have better root morphology (length, mass, surface area, diameter, and volume) under imposition of salt treatments (Sheng *et al.* 2009).

The better performance of photosystem-II and stomatal conductance, and more accumulation of antioxidants of maize plants inoculated with native AMF sp. jointly contributed to enhance salt tolerance (Estrada *et al.* 2013). Chlorophyll content, photosynthesis and leaf water content of peanut plants were found to increase under salinity stress by the inoculation with mycorrhizal fungi (*Glomus mosseae*) (Al-Khalil 2010). AMF inoculated citrus seedlings showed more contents of ascorbate, soluble protein and glutathione and greater activity of catalase under both saline and non-saline conditions and thus imparted better antioxidant defense system in plants (Wu *et al.* 2010b). On a similar note another group of worker also evidenced that AMF may protect tomato plants against salinity by alleviating the salt induced oxidative stress by increasing of superoxide dismutase, catalase, peroxidase and ascorbate peroxidase and reduced malon-di-aldehyde (MDA) content (Latef and Chaoxing, 2011).

Biochemical alterations

Plant tolerance to salinity is a complex trait to which plants have evolved responses involving biochemical and molecular mechanisms. Additional plant responses can include selective build-up or exclusion of salt ions, maintenance of photosynthesis at values adequate for plant growth, changes in membrane structure and synthesis of phytohormones (Turkan and Demiral 2009)

AMF are reported to improve tolerance and/or resistance of plants against salinity by increasing accumulation of these compounds.

Proline, betaine accumulation in plant is the most studied modifications following salt exposure.



Proline and betaine accumulation has been found to increase when the *Glycine max* is colonized by *Glomus etunicatum* (Sharifi *et al.* 2007) and *Phragmites australis* colonized by *Glomus fasciculatum* (Al-Garni 2006).

Free polyamines mainly putrescine, spermidine and spermine are recognized as a key role in plant responses to a wide array of environmental stressors such as salinity, osmolarity and antioxidative stress (Evelin *et al.* 2009). They are essential in regulating root development under salt exposure (Couee *et al.* 2004). Level of free polyamine pools is declined under salt stress condition; nevertheless, the inoculation of host plants with AMF increases free polyamine concentrations (Sannazzaro *et al.* 2007).

Abscisic acid (ABA), a phytohormone, is well known for its signalling role in the regulation of plant growth and development in response to the salinity stress by closing the stomata to reduce water loss and inducing the expression of stress-related genes that reduces the adverse effect it has caused (Evelin *et al.* 2009).

Another plant tolerance mechanism involves accumulation of soluble sugars that lowered down the osmotic potential of the plant (Thanna and Nawar 1994). Plants (*Phragmites australis*, *Glycine max*) colonized by AMF (*Glomus fasciculatum*, *Glomus intraradices*) are reported to have higher level of soluble sugars than those non-mycorrhizal plants (Porcel and Ruiz-Lozano 2004; Al-Garni 2006). Moreover, Trehalose, is a non-reducing disaccharide acts as the prime storage carbohydrate in AMF. It functions as abiotic stress protectant that stabilizes dehydrated enzymes and membranes, safeguards biological structures from desiccation damage. Although, presence of trehalose is very infrequent in higher plants, but it gets induced by AMF colonization of plant roots, in turn protects plants against salt stress (Hoekstra *et al.* 1992; Schubert *et al.* 1992).

Plants with high concentrations of antioxidants have been perceived to impart greater resistance against oxidative damage (Jiang and Zhang 2002). Most common antioxidant enzymes are superoxide dismutase, catalase, dehydroascorbate reductase, glutathione reductase and different peroxidase. Plants having their roots colonized by AMF are reported to abate salinity stress through augmenting

the activities of antioxidant enzymes (Ghorbanli *et al.* 2004; Zhong Qun *et al.* 2007).

Chlorophyll content in leaves is another parameter which could differentiate between mycorrhizal and non-mycorrhizal plants exposed to salt stress. AMF colonization is reported to reduce salt interference in chlorophyll synthesis (Giri and Mukerji 2004; Zuccarini 2007; Colla *et al.* 2008; Sheng *et al.* 2008) as well as increases chlorophyll content in *Capsicum annuum*, *Glycine max*, *Cyamopsis tetragonoloba* and *Zea mays* (Beltrano *et al.* 2013; Datta and Kulkarni 2014) Furthermore, the antagonistic effect of Na⁺ ion on Mg²⁺ uptake is counter balanced and suppressed (Giri *et al.* 2003) in the presence of mycorrhiza. Chlorophyll fluorescence, a measure of photosynthetic efficiency is calculated as the ratio between variable and maximum fluorescence (Fv/Fm) which indicates capacity of the primary photochemistry of PSII (Sheng *et al.* 2008; Zuccarini and Okurowska 2008). The ratio Fv:Fm is very sensitive to salt stress. Nevertheless, mycorrhiza helps plants to maintain significantly higher Fv:Fm ratio thus maintaining the photosynthetic efficiency under saline stress condition (Sheng *et al.* 2008; Zuccarini and Okurowska 2008).

Various studies have also documented that AMF symbiosis helps plants to alleviate salt- or water-deficit stresses by enhancing the activities of antioxidant enzymes such as superoxide dismutase, catalase, ascorbate peroxidase, glutathione reductase, dehydroascorbate reductase, monodehydroascorbate reductase, guaiacol peroxidase, oxidized glutathione, glutathione peroxidase (Zhong *et al.* 2007; Garg and Manchanda 2009; Talaat and Shawky 2011). Thus, mycorrhizal plants possess enhanced activity of several antioxidant enzymes.

Improved nutrient uptake

Facilitating the nutrient uptake for plants by use mycorrhiza has repeatedly been highlighted by the researchers throughout the world. Apart from primary soil nutrients (N, P, K), AMF proved its efficiency to absorb Mg, Ca, Cu, Zn, Fe, Ni, Cd through plant roots. It is thus often been considered that uptake and transport of nutrients from soil is the primary function of mycorrhizal fungi associated with plant roots (reviewed in Quilambo 2003). The fungal hyphal network is ideally positioned to efficiently take up nutrients and water from soil, but



only a few fungal transporters are involved in this process, including those that transport phosphate, ammonium and zinc. Because of diffusion is too slow the nutrients are moved in a packaged form between the extra-radicle and the intra-radicle mycelium. Possible mechanisms of nutrient uptake are:

- (a) Better absorption of nutrients: The mycorrhizal plants absorb the nutrients more efficiently compared to non-mycorrhizal ones as AMF changes root space geometry and increases root surface area, acts as absorbing surface, increases the efficiency of roots to absorb nutrients and also enhances the life span of roots keeping them active for greater period.
- (b) Increasing nutrient availability: Some AMF synthesize phosphatases which increase mineralization of organic phosphate and increase phosphate availability while few AMF produce organic acids which increase pH and in turn solubility and availability of phosphate.
- (c) Increasing availability: AMF may also solubilize some unavailable form of mineral phosphate to available form enhancing its availability.

Phosphorus absorption has often been easier under mycorrhizal inoculation; even under saline soil the P uptake was found higher (Tian *et al.* 2004; Sharifi *et al.* 2007; Al-Khaliel 2010). P, being a poorly mobile nutrient as PO_4^{3-} , when show a positive influence in presence of AMF towards absorption in plant roots, and that also under problem (saline) soil, demands a special mention. Despite P, N and K uptake were also found improved by association of AMF (Rabie and Almadini 2005; Al-Khaliel 2010). Garg and Manchanda (2008) and Giri and Mukerji (2004) both reported a higher N uptake in presence of *Glomus* sp. under soil salinity by *Cajanus cajan* and *Sesbania* sp. respectively. Enhancement of potassium uptake under salt-stressed soil was also found for Soybean plants (Sharifi *et al.* 2007).

A selective uptake of nutrients is also sometimes mentioned by AMF. Balancing the K^+/Na^+ ratio in plant tissues is a major concern to avoid the deleterious effects of soil salinity. AMF was also found to interfere in the increased uptake of K with concomitant decreased uptake of Na by plant

roots (Zuccarini and Okurowska 2008). Ca^{+2} and Mg^{+2} were also found to absorb more by plant roots with mycorrhizal association despite of soil salinity (Yano-Melo *et al.* 2003; Sharifi *et al.* 2007; Giri and Mukerji 2004).

Negative effect of salinity can also be alleviated by AM-fungi due to reduced uptake of Ca and K and increase in levels of P, Zn and Cu in leaves (Ezz and Nawar 1993). AM fungi improved plant tolerance to salinity through better host plant nutrition, and maintained higher K^+/Na^+ ratio in plant tissues (Estrada *et al.* 2013).

This sort of beneficial activities by AMF is often enhances the nutrient use efficiency of plants in marginal or degraded soils, depleted with the essential nutrients. This may further aggravate the soil fertility and productivity.

Potential genera of AMF

Arbuscular mycorrhizal fungi have been reported to occur naturally in saline environments. The most commonly observed genera in saline soils is *Glomus* spp. belonging to family Glomeromycota, however, the ecological specificity has not yet been demonstrated. Studies of early dates employed only morphological characterization to identify the AMF spores. Nevertheless, fatty acid methyl ester (FAME) study, use of molecular techniques such as polymerase chain reaction (PCR) and restriction fragment length polymorphism (RFLP) are aiding better and more accurate enumeration of the AM fungal diversity in saline soil area. The dominant genera of AM fungi having potential of salt stress amelioration are mainly *Glomus intraradices*, *G. versiform*, *G. Etunicatum* (Aliasgharzadeh *et al.* 2001; Porras-Soriano *et al.* 2009) *G. fasciculatum*, *G. macrocarpum*, *G. geosporum*, *G. coronatum* (Giri and Mukerji 2004; Giri *et al.* 2007), *Gigaspora gigantean*, *G. margarita* (Sambandan 2014), *Acaulospora*, *Archaeospora* (Wang *et al.* 2004), *Funneliformis mosseae*, *F. geosporum*, *F. coronatum*, *Rhizofagus fasciculatus* (Bencherif *et al.* 2015).

Crop productivity enhancement through AM Fungi intervention in salt affected soils

AMF with their favourable symbiotic relationship with plants could also play a crucial role in enhancing crop productivity in salt affected soil.

Table 1: Potential genera of arbuscularmycorrhizal fungi reported for combating salinity stress

Sl. No	Potential Genera	Plant growth benefit
1	<i>Glomus intraradices</i> , <i>Glomus mosseae</i> <i>Glomus mosseae</i>	Improved growth, yield and quality of fruits in Pumpkin. Enhanced root and shoot growth and lesser biomass reduction in olive plant. In maize plants imparted better root morphology (length, mass, surface area, diameter and volume).
3	<i>Glomus mosseae</i> , <i>Glomus fasciculatum</i>	Increased Proline content in Maize, Soybean
4	<i>Glomus mosseae</i> , <i>Glomus fasciculatum</i> , <i>Glomus clarum</i>	Increased soluble carbohydrate content in Green gram, Soybean.
5.	<i>Glomus intraradice</i> , <i>Glomus etunicatum</i>	Increased Abscisic acid in Lettuce and Tomato.
6.	<i>Glomus mosseae</i> , <i>Glomus versiforme</i> , <i>Glomus intraradices</i> , <i>Glomus etunicatum</i>	Increased antioxidant enzyme activity in tomato and cowpea.

With the help of their extra-matrical hyphae they increase the absorptive surface area of the roots of the host plants leading to more accessibility and absorption of relatively immobile elements in soil such as P, Cu and Zn (Menge and Timmer 1982). Till date, only few studies have reported the role of AMF in direct crop productivity enhancement. Tripartite association among AMF, *Bradyrhizobium* and mungbean (*Vigna radiate* L.) greatly helped in establishment and cultivation of the crop in the saline soils of West Bengal, India (Singh *et al.*, 2011). Daei *et al.* (2009) studied the influence of *Glomus etunicatum*, *G. mosseae*, *G. intraradices* on enhancing wheat growth under salinity, where the electrical conductivity of the irrigation water was as high as 13.87 dS/m. The increase in grain yield ranged from 4.5 to 38.3% mainly due to *Glomus etunicatum* and *G. intraradices*, respectively. *Glomus etunicatum* appeared to be the best performer among three species. This reflected the great significance of choosing the right combination of AM species and host plant to exploit the symbiosis benefit on crop productivity under salinity condition. Similarly, a study on influence of combined application of mycorrhizal fungi and compost reported improved berseem yield owing to increased uptake of nutrients (P, Mn, Zn, Fe, Cu) in alkaline calcareous soil (Jan *et al.* 2014).

CONCLUSION

Role of AMF in abating salt stress on crop growth is still unexplored. Thus, ample research opportunity is there to screen out the suitable location-specific as well as crop-specific AM fungal strains. The most efficient AM fungal strain needs to be exploited for

enhanced productivity under salinity-prone areas. In this connection, the emerging strains can be studied for identifying their various morphological, and biochemical effects on plants. The newly identified strains can also be tested for facilitated uptake of single or multiple nutrients ions through plant cells. Moreover, studies on AMF diversity as well as signaling mechanism, prerequisite for host plant colonization by AM fungal species, are also necessary for understanding the ecological function played by AMF under wide range of salt affected condition. Further the potentiality of AMF can be tested for multiple abiotic soil stresses along with salinity.

REFERENCES

- Akbarimoghaddam, H., Galavi, M., Ghanbari, A. and Panjehkeh, N. 2011. Salinity effects on seed germination and seedling growth of bread wheat cultivars. *Trakia journal of Sciences*, **9** (1): 43–50.
- Al-Garni, S.M.S. 2006. Increasing NaCl-salt tolerance of a halophytic plant *Phragmites australis* by mycorrhizal symbiosis. *American-Eurasian Journal of Agricultural and Environmental Science*, **1**: 119–126.
- Aliasgharzadeh, N., Saleh Rastin, N., Towfighi, H. and Alizadeh, A. 2001. Occurrence of arbuscular mycorrhizal fungi in saline soils of the Tabriz Plain of Iran in relation to some physical and chemical properties of soil. *Mycorrhiza*, **11**: 119–122.
- Al-Karaki, G.N. 2000. Growth of mycorrhizal tomato and mineral acquisition under salt stress. *Mycorrhiza*, **10**: 51–54.
- Al-Karaki, G.N., Hammad, R. and Rusan, M. 2001. Response of two tomato cultivars differing in salt tolerance to inoculation with mycorrhizal fungi under salt stress. *Mycorrhiza*, **11**(1): 43–47.
- Al-Khaliel, A.S. 2010. Effect of salinity stress on mycorrhizal association and growth response of peanut infected by *Glomus mosseae*. *Plant Soil and Environment*, **56**(7): 318–324.



- Allen, E.B. and Cunningham, G.L. 1983. Effects of vesicular-arbuscularmycorrhizae on *Distichlis spicata* under three salinity levels. *New Phytologist*, **93**: 227–236.
- Ashraf, M. 2004. Some important physiological selection criteria for salt tolerance in plants. *Flora*, **199**: 361–376.
- Bano, A. and Fatima, M. 2009. Salt tolerance in *Zea mays* (L.) following inoculation with *Rhizobium* and *Pseudomonas*. *Biology and Fertility of Soils*, **45**: 405–413.
- Beltrano, J. and Ronco, M.G. 2008. Improved tolerance of wheat plants (*Triticum aestivum* L.) to drought stress and re-watering by the arbuscular mycorrhizal fungus *Glomus claroideum*: Effect on growth and cell membrane stability. *Brazilian Journal of Plant Physiology*, **20**(1): 29–37.
- Beltrano, J., Ruscitti, M., Arango, M.C. and Ronco, M. 2013. Effects of arbuscular mycorrhiza inoculation on plant growth, biological and physiological parameters and mineral nutrition in pepper grown under different salinity and P levels. *Journal of Soil Science and Plant Nutrition*, **13**: 123–141.
- Bhaduri, D., Meena, H.N. and Chakraborty, K. 2016. Variation in phosphorus accumulation in groundnut cultivars as influenced by water salinity. *Legume Research: An International Journal*, **39**(2): 215–220.
- Blaylock, A.D. 1994. Soil salinity, salt tolerance and growth potential of horticultural and landscape plants. Cooperative Extension Service, University of Wyoming, Department of Plant, Soil and Insect Sciences, College of Agriculture, Laramie, Wyoming.
- Borkowska, B. 2002. Growth and photosynthetic activity of micropropagated micro propagated strawberry plants inoculated with endomycorrhizal fungi (AMF) and growing under drought stress. *Acta Physiologiae Plantarum*, **24**(4): 365–370.
- Calvet, C., Pinochet, J., Camprubí, A. and Fernández, C. 1995. Increased tolerance to the root-lesion nematode *Pratylenchus vulnus* in mycorrhizal micro propagated BA-29 quince rootstock. *Mycorrhiza*, **5**(4): 253–258.
- Chakraborty, K., Singh, A.L., Bhaduri, D. and Sairam, R.K. 2013. Mechanism of salinity stress tolerance in crop plants and recent developments. In: *Advances in plant physiology* vol. 14 (Ed. A. Hemantaranjan) Scientific Publishers (India), Jodhpur. pp.466–496.
- Chakraborty, K., Sairam, R.K. and Bhaduri, D. 2015. Effects of Different Levels of Soil Salinity on Yield Attributes, Accumulation of Nitrogen, and Micronutrients in *Brassica* Spp. *Journal of Plant Nutrition*, **39**(7): 1026–1037.
- Chakraborty, K., Bishi, S.K., Goswami, N., Singh, A.L. and Zala, P.V. 2016. Differential fine-regulation of enzyme driven ROS detoxification network imparts salt tolerance in contrasting peanut genotypes. *Environmental and Experimental Botany*, **128**: 79–90.
- Chinnusamy, V., Zhu, J. and; Zhu, Jian-Kang. 2006. Gene regulation during cold acclimation in plants. *Physiol. Plant.*, **126**(1): 52–61.
- Colla, G., Roupheal, Y., Cardarelli, M., Tullio, M., Rivera, C.M. and Rea, E. 2008. Alleviation of salt stress by arbuscular mycorrhizal in zucchini plants grown at low and high phosphorus concentration. *Biology and Fertility of Soils*, **44**: 501–509.
- Coué, I., Hummel, I., Sulmon, C., Gowsbet, G. and El Armani, A. 2004. Involvement of polyamines in root development. *Plant Cell, Tissue and Organ Culture*, **76**: 1–10.
- Daei, G., Ardekani, M.R., Rejali, F., Teimuri, S. and Miransari, M. 2009. Alleviation of salinity stress on wheat yield, yield components, and nutrient uptake using arbuscular mycorrhizal fungi under field conditions. *Journal of Plant Physiology*, **166**(6): 617–625.
- Datta, P. and Kulkarni, M. 2014. Arbuscular mycorrhizal colonization enhances biochemical status in and mitigates adverse salt effect on two legumes. *Notulae Scientia Biologicae*. **6**: 381–393.
- Dixon, R. K., Garg, V.K. and Rao, M.V. 1993. Inoculation of *Leucaena* and *Prosopis* seedlings with *Glomus* and *Rhizobium* species in saline soil: rhizosphere relations and seedlings growth. *Arid Soil Res Rehabil*, **7**: 133–144.
- Estrada, B., Aroca, R., Barea, J.M. and Ruiz-Lozano, J.M. 2013. Native arbuscular mycorrhizal fungi isolated from a saline habitat improved maize antioxidant systems and plant tolerance to salinity. *Plant science*, **201**: 42–51.
- Estrada, B., Aroca, R., Maathuis, F.J.M., Barea, J.M. and Ruiz-Lozano, J.M. 2013. Arbuscular mycorrhizal fungi native from a mediterranean saline area enhance maize tolerance to salinity through improved ion homeostasis. *Plant Cell and Environment*, DOI/10.1111/pce.12082.
- Evelin, H., Kapoor, R. and Giri, B. 2009. Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. *Annals of Botany*, **104**: 1263–1280.
- Ezz, T. and Nawar, A. 1993. Salinity and mycorrhizal infection on growth and mineral nutrition of sour orange seedlings. *Alexandria Journal of Agricultural Research*, **38**: 439–457.
- Feng, G., Zhang, F.S., Li, X., Tian, C.Y.; Tang, C. and Rengel, Z. 2002. Improved tolerance of maize plants to salt stress by arbuscular mycorrhiza is related to higher accumulation of soluble sugars in roots. *Mycorrhiza*, **12**: 185–190.
- Garg, N. and Manchanda, G. 2008. Effect of arbuscular mycorrhizal inoculation of salt-induced nodule senescence in *Cajanus cajan* (pigeonpea). *Journal of Plant Growth Regulators*, **27**: 115–124.
- Garg, N. and Manchanda, G. 2009. Role of arbuscular mycorrhizae in the alleviation of ionic, osmotic and oxidative stresses induced by salinity in *Cajanus cajan* (L.) millsp. (pigeon pea). *Journal of Agronomy and Crop Science*, **195**: 110–123.
- Giri, B. and Mukerji, K.G. 2004. Mycorrhizal inoculant alleviates salt stress in *Sesbania aegyptiaca* and *Sesbania grandiflora* under field conditions: evidence for reduced sodium and improved magnesium uptake. *Mycorrhiza*, **14**: 307–312.
- Giri, B., Kapoor, R. and Mukerji, K.G. 2007. Improved tolerance of *Acacia nilotica* to salt stress by arbuscular mycorrhiza, *Glomus fasciculatum*, may be partly related to elevated K⁺/Na⁺ ratios in root and shoot tissues. *Microbial Ecology*, **54**: 753–760.

- Giri, B.; Kapoor, R. and Mukerji, K.G. 2003. Influence of arbuscularmycorrhizalfungi and salinity on growth, biomass and mineral nutrition of *Acacia auriculiformis*. *Biology and Fertility of Soils*, **38**: 170–175.
- Hajiboland, R., Aliasgharzadeh, N., Laiegh, S.F. and Poschenrieder, C. 2010. Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum lycopersicum* L.) plants. *Plant and Soil*, **331**: 313–327.
- Heijden, J.N., Klironomos, M., Ursic, P. et al. 1998. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature*, **396**: 69–72.
- Hirsch, A.M. and Kapulnik, Y. 1998. Signal transduction pathways in Mycorrhizal associations: Comparisons with the *Rhizobium*–Legume symbiosis. *Fungal Genetics and Biology*, **23**: 205–212.
- Ho, I. 1987. Vesicular-arbuscularmycorrhizae of halophytic grasses in the Alvord desert of Oregon. *Northwest Science*, **61**: 148–151.
- Hodge, A. 2000. Microbial ecology of the arbuscular mycorrhiza. *Microbiol Ecology*, **32**: 91–96.
- Hoekstra, F.A., Crow, J.H., Crowe, L.M., Van Roekel, T. and Vermeer, T. 1992. Do phospholipids and sucrose determine membrane phase transitions in dehydrating pollen species? *Plant, Cell and Environment*, **15**: 601–606.
- Jan, B., Ali, A., Wahid, F., Noor, S., Shah, M., Khan, A., Khan, F. 2014. Effect of Arbuscular Mycorrhiza Fungal Inoculation with Compost on Yield and Phosphorous Uptake of Berseem in Alkaline Calcareous Soil. *American Journal of Plant Sciences*, **5**: 1359-1369.
- Javid, M.G.; Sorooshzadeh, A., Moradi, F., SanavySeyed, A. M.M. and Allahdadi, I. 2011. The role of phytohormones in alleviating salt stress in crop plants. *Australian Journal of Crop Science*, **5**(6): 726–734.
- Jiang, M. and Zhang, J. 2002. Water stress-induced abscisic acid accumulation triggers the increased generation of reactive oxygen species and p-regulates the activities of antioxidant enzymes in maize-leaves. *Journal of Experimental Botany*, **53**: 2401–2410.
- Juniper, S. and Abbott, L.K. 2006. Soil salinity delays germination and limits growth of hyphae from propagules of arbuscular mycorrhizal fungi. *Mycorrhiza*, **16**(5): 371–379.
- Latef, A.A.H.A. and Chaoping, H. 2011. Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. *Scientia Horticulturae*, **127**(3): 228-233.
- Mathur, N., Singh, J., Bohra, S. and Vyas, A. 2007. Arbuscular mycorrhizal status of medicinal halophytes in saline areas of Indian Thar Desert. *International Journal of Soil Science*, **2**: 119–127.
- McCue, K.F. and Hanson, A.D. 1992. Salt-inducible betaine aldehyde dehydrogenase from sugar beet: cDNA cloning and expression. *Plant Molecular Biology*, **18**(1): 1-11.
- Menge, J.A. and Timmer, L.W. 1982. Procedure for Inoculation of Plants Vesicular-Arbuscular Mycorrhizal in Laboratory, Green House and Field. In: Schenck, N.C., Ed., *Methods and Principles of Mycorrhizal Research*, American Phytopathological Society, St. Paul.
- Munns, R. and Tester, M. 2008. Mechanisms of salinity tolerance. *Annual Review in Plant Biology*, **59**: 651–681.
- Netondo, G.W., Onyango, J.C. and Beck, E. 2004. Sorghum and salinity: II. Gas exchange and chlorophyll fluorescence of sorghum under salt stress. *Crop Science*, **44**: 806–811.
- Pal, S., Singh, H.B., Rai, A. and Farooqui, A. 2017. Diversity of Arbuscular Mycorrhiza Associated with Long Term Wastewater Irrigation in the Peri-urban Soil of Varanasi. *International Journal of Agriculture, Environment & Biotechnology*, **10**: 779-784.
- Pal, S., Singh, H.B., Rai, A. and Rakshit, A. 2013. Evaluation of Different Medium for Producing on farm Arbuscular Mycorrhizal Inoculum. *International Journal of Agriculture, Environment & Biotechnology*, **6**: 557-562
- Plenchette, C. and Dupponois, R. 2005. Growth response of the salt brush *Atriplex nummularia* L. to inoculation with the arbuscular mycorrhizal fungus *Glomus intraradices*. *Journal of Arid Environments*, **61**: 535–540.
- Porcel, R. and Ruiz-Lozano, J.M. 2004. Arbuscular mycorrhizal influence on leaf water potential, solute accumulation and oxidative stress in soybean plants subjected to drought stress. *Journal of Experimental Botany*, **55**: 1743–1750.
- Porcel, R., Aroca, R. and Juan Ruiz-Lozano, J.M. 2012. Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. *Agronomy for Sustainable Development*, **32**(1): 181-200.
- Porrás-Soriano, A., Soriano-Martin, M.L., Porrás-Piedra, A. and Azcón, R. 2009. Arbuscularmycorrhizal fungi increased growth, nutrient uptake and tolerance to salinity in olive trees under nursery conditions. *Journal of Plant Physiology*, **166**(13): 1350-1359.
- Porrás-Soriano, A., Soriano-Martin, M.L., Porrás-Piedra, A. and Azcón, R. 2009. Arbuscular mycorrhizal fungi increased growth, nutrient uptake and tolerance to salinity in olive trees under nursery conditions. *Journal of plant physiology*, **166**(13): 1350-1359.
- Quilambo, O.A. 2004. The vesicular-arbuscular mycorrhizal symbiosis. *African Journal of Biotechnology*, **2**(12): 539-546.
- Rabie, G.H. and Almadini, A.M. 2005. Role of bio-inoculants in development of salt-tolerance of *Vicia faba* plants under salinity stress. *African Journal of Biotechnology*, **4**: 210–222.
- Raverkar, K.P. and Bhattacharya, S. 2014. Arbuscular Mycorrhizae: Status and Potential. In: Bioresources for Sustainable Plant Nutrient Management (Eds: Ramesh Chandra and K P Raverkar) Satish Serial Publishing House, New Delhi, DOI: 10.13140/RG.2.1.2532.5848.
- Rosendahl, C.N. and Rosendahl, S. 1991. Influence of vesicular-arbuscular mycorrhizal fungi (*Glomus* spp.) on the response of cucumber (*Cucumis sativus* L.) to salt stress. *Environmental and Experimental Botany*, **31**: 313–318.



- Ruiz-Lozano, J.M., Azcon, R. and Gomez, M. 1996. Alleviation of salt stress by arbuscular-mycorrhizal *Glomus* species in *Lactuca sativa* plants. *Physiologia Plantarum*, **98**: 767–772.
- Sairam, R.K. and Tyagi, A. 2004. Physiology and molecular biology of salinity stress tolerance in plants. *Curr. Sci.*, **86**: 407–421.
- Sannazzaro, A.I., Echeverria, M., Alberto, E.O., Ruiz, O.A. and Menéndez, A.B. 2007. Modulation of polyamine balance in *Lotus glaber* by salinity and arbuscular mycorrhiza. *Plant Physiology and Biochemistry*, **45**: 39–46.
- Schubert, A., Wyss, P. and Wiekman, A. 1992. Occurrence of trehalose in vesicular-arbuscularmycorrhizal fungi and in mycorrhizal roots. *Journal of Plant Physiology*, **140**: 41–45.
- Seckin, B., Sekmen, A.H. and Turkan, I. 2009. An enhancing effect of exogenous mannitol on the antioxidant enzyme activities in roots of wheat under salt stress. *Journal of Plant Growth Regulation*, **28**: 12–20.
- Sharifi, M., Ghorbanli, M. and Ebrahimzadeh, H. 2007. Improved growth of salinity-stressed soybean after inoculation with pre-treated mycorrhizal fungi. *Journal of Plant Physiology*, **164**: 1144–1151.
- Sheng, M., Tang, M., Chen, H., Yang, B., Zhang, F. and Huang, Y. 2009. Influence of arbuscularmycorrhizae on the root system of maize plants under salt stress. *Canadian Journal of Microbiology*, **55**(7): 879–886.
- Singh, N., Samajapati, N. and Paul, A.K. 2011. Dual inoculation of salt tolerant *Bradyrhizobium* and *Glomus mosseae* for improvement of *Vigna radiata* L. cultivation in saline areas of West Bengal, India. *Agricultural Sciences*, **2**(4): 413–423.
- Smith, F.A., Jakobsen, I. and Smith, S.E. 2000. Spatial differences in acquisition of soil phosphate between two arbuscular mycorrhizal fungi in symbiosis with *Medicago truncatula*. *New Phytologist*, **147**: 357–366.
- Smith, S.E. and Read, D.J. (eds) 2008. *Mycorrhizal symbiosis*. Academic Press, Inc., San Diego, USA.
- Tabur, S. and Demir, K. 2010. Role of some growth regulators on cytogetic activity of barley under salt stress. *Plant Growth Regulation*, **60**: 99–104.
- Talaat, N.B. and Shawky, B.T. 2011. Influence of arbuscular mycorrhizae on yield, nutrients, organic solutes, and antioxidant enzymes of two wheat cultivars under salt stress. *Journal of Plant Nutrition and Soil Science*, **174**: 283–291.
- Thanna, E. and Nawar, A. 1994. Salinity and mycorrhizal association in relation to carbohydrate status, leaf chlorophyll and activity of peroxidase and polyphenol oxidase enzymes in sour orange seedlings. *Alexandria Journal of Agricultural Research*, **39**: 263–280.
- Tian, C.Y., Feng, G., Li, X.L. and Zhang, F.S. 2004. Different effects of arbuscular mycorrhizal fungal isolates from saline or non-saline on salinity tolerance of plants. *Applied Soil Ecology*, **26**: 143–148.
- Turkan, I. and Demiral, T. 2009. Recent developments in understanding salinity tolerance. *Environmental and Experimental Botany*, **67**: 2–9.
- Wang, F.Y., Liu, R.J., Lin, X.G. and Zhou, J.M. 2004. Arbuscular mycorrhizal status of wild plants in saline-alkaline soils of the Yellow River Delta. *Mycorrhiza*, **14**: 133–137.
- Wu, Q.S., Zou, Y.N. and He, X.H. 2010a. Contributions of arbuscular mycorrhizal fungi to growth, photosynthesis, root morphology and ionic balance of citrus seedlings under salt stress. *Acta Physiologiae Plantarum*, **32**(2): 297–304.
- Wu, Q.S., Zou, Y.N., Liu, W., Ye, X.F., Zai, H.F. and Zhao, L. J. 2010b. Alleviation of salt stress in citrus seedlings inoculated with mycorrhiza: changes in leaf antioxidant defense systems. *Plant Soil and Environment*, **56**(10): 470–475.
- Yano-Melo, A.M., Saggin, O.J. and Maia, L.C. 2003. Tolerance of mycorrhized banana (*Musa* sp. cv. Pacovan) plantlets to saline stress. *Agriculture, Ecosystems and Environment*, **95**: 343–348.
- Yadav, R.S., Mahatma, M.K., Thirumalaisamy, P.P., Meena, H.N., Bhaduri, D., Arora, S. and Panwar, J. 2017. Arbuscular mycorrhizal fungi (AMF) for sustainable soil and plant health in salt-affected soils. In *Bioremediation of Salt Affected Soils: An Indian Perspective* (pp. 133–156). Springer, Cham.
- Zhong Qun, H., Chao Xing, H., Zhibin, Z., Zhirong, Z. and Huai Song, W. 2007. Changes in antioxidative enzymes and cell membrane osmosis in tomato colonized by arbuscular mycorrhizae under NaCl stress. *Colloids and Surfaces B: Biointerfaces*, **59**: 128–133.
- Zhong Qun, H., Chao, X.H., Zhibin, Z., Zhirong, Z. and Huai, S.W. 2007. Changes in antioxidative enzymes and cell membrane osmosis in tomato colonized by arbuscular mycorrhizae under NaCl stress. *Colloids and Surfaces B: Biointerfaces*, **59**: 128–133.
- Zuccarini, P. 2007. Mycorrhizal infection ameliorates chlorophyll content and nutrient uptake of lettuce exposed to saline irrigation. *Plant, Soil and Environment*, **53**: 283–289.
- Zuccarini, P. and Okurowska, P. 2008. Effects of mycorrhizal colonization and fertilization on growth and photosynthesis of sweet basil under salt stress. *Journal of Plant Nutrition*, **31**: 497–513.