

# Serological evidence of Avian Pneumovirus infection in broiler and layer chickens in Grenada, West Indies

**Keshaw Tiwari, Alfred Chikweto, Muhammed Iqbal Bhaiyat, Claude DeAllie, Graeme Stratton and Ravindra Sharma\***

Pathobiology Academic Program, School of Veterinary Medicine  
St George's University, Grenada, WEST INDIES

\*Corresponding Author: R Sharma; Email: rsharma@sgu.edu

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## ABSTRACT

A serological survey was conducted to detect avian pneumovirus (APV) antibodies in commercial poultry birds in Grenada using a commercially available enzyme-linked immunosorbent assay (ELISA) kit. Sera were collected from 226 layers and 233 broilers. Age of the layers ranged from 12 to 18 months while that of the broilers ranged from 6 to 7 weeks. One hundred forty of the layers (61.9%) and 74 of the broilers (31.8%) were positive for APV antibodies. Chickens are not vaccinated for APV in Grenada and these results indicate that commercial poultry birds are exposed to this important poultry pathogen. This is the first report of serologic evidence of APV in Grenada and the Eastern Caribbean region.

**Keywords:** avian pneumovirus, chicken, Grenada, serology

Avian pneumovirus (APV), a metapneumovirus of the family paramyxoviridae, causes turkey rhinotracheitis (TRT), swollen head syndrome (SHS) and avian rhinotracheitis (ART). APV was first detected in turkeys in South Africa in the late 1970s (Buys and du Preez 1980). Later it was reported in chickens in South Africa (Morley and Thomson, 1984). By 1993, Alexander described TRT in Israel, France and Great Britain. Serological evidence of APV is now available from many countries of the world (Cook 1997). There is no published information on APV in Grenada and other countries of the East Caribbean region. The present study was conducted to find out the status of APV infection in commercial chickens in Grenada.

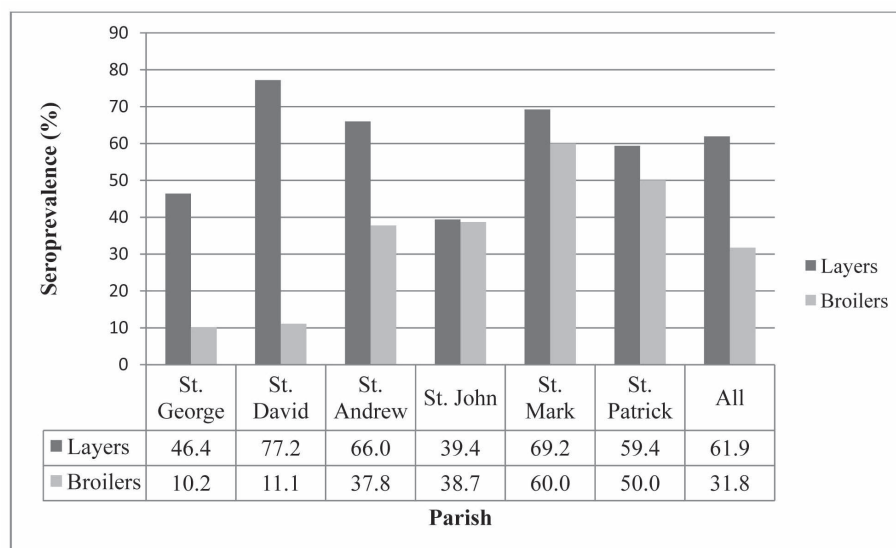
## MATERIALS AND METHODS

A total of 459 blood samples were collected from commercial poultry flocks in various Parishes of mainland Grenada. Birds from layer flocks (n=226 birds) and broiler flocks (n=233 birds) were analyzed. The age of the layers varied between 12 and 18 months and the broilers were between 6 and 7 weeks of age. Blood samples from layers were obtained by brachial venipuncture. Blood from broilers

was collected during slaughter. A minimum of 25 blood samples were taken separately from layer and broiler flocks in each Parish. Sera were stored at -20°C until tested for APV antibodies using a commercial ELISA (IDEXX APV) kit following the instructions of the manufacturers.

## RESULTS

The survey revealed that 61.9% layers and 31.8% broilers in Grenada were serologically positive for APV (Figure 1). In the Parish of St John the seroprevalence was almost equal in layers and broilers, whereas a higher rate of seropositivity was observed in layers in the other five Parishes (St. Mark, St. Patrick, St David and St. Andrew) as depicted in Figure 1.



## DISCUSSION

A higher rate of seropositivity for APV in layer chickens observed in Grenada is in agreement with previous reports from other part of the world (Gough 2003; Owoade *et al.* 2006), suggesting that the virus in layer flocks is being transmitted between age group of flocks either by contact between flocks or by contamination of infrastructure. In Grenada, layers are usually maintained as multi age flocks. Transmission between flocks could be avoided by adopting an “all-in, all-out” replacement system of management and regular periodic decontamination. A lower rate of seropositivity for APV in broilers is expected since most broiler flocks in Grenada are managed as an “all-in, all-out” system.

In broilers, APV has been associated with clinical signs of swollen head syndrome (SHS). Layers infected with APV in the early phase of lay-do not reach peak production, whereas layers in their late phase of lay suffer a drop in egg production

(Paul McMullin 1998, Cook 2000). No clinical signs were reported in the broiler and layer flocks at the time of blood sample collection in this investigation. This is in agreement with previous reports from other parts of the world. APV has been isolated from chicken flocks without clinical signs (Johns *et al.*, 1991) and chicken flocks free of clinical signs may have antibodies for APV (Cook *et al.*, 1988; Pattison *et al.*, 1989; Hafez and Lohren 1990).

Various strains of APV have been identified. Using serological and molecular techniques, APV has been grouped into three serotypes; A, B and C. The APV isolates from the United States has been grouped under serotype C (Cook *et al.*, 1999; Seal 2000). Vaccines prepared from serotype A may confer immunity against serotype B (Cook *et al.*, 1995). Chickens are not vaccinated for APV in Grenada and the seropositivity for APV indicates that commercial chickens in Grenada are exposed to this important poultry pathogen. APV may cause serious economic losses in turkeys and chickens especially in the presence of concurrent bacterial and viral infections. Therefore, further studies to isolate and identify the strain of APV are warranted. Results of such a study would be strategic for institution of preventive measures against APV infection.

## REFERENCES

- Alexander, D.J. 1993. Pneumovirus (turkey rhinotracheitis and Swollen head syndrome of chickens) I. J. B. McFerran and M. S. McNulty (eds). Virus infections of birds. Elsevier Science Publishers: B. V. 375-382.
- Buys, S.B. and du Preez, J.H. 1980. A preliminary report on the isolation of a virus causing sinusitis in turkeys in South Africa and attempts to attenuate the virus. *Turkeys* (June), **36**: 56.
- Cook, J.K.A., Dolby, C.A., Southee, D.J. and Mockett, A.P.A. 1988. Demonstration of antibodies to turkey rhinotracheitis virus in serum from commercially reared flocks of chickens. *Avian Pathology*, **17**: 403-410.
- Cook, J. K. A., Huggins, M. B., Woods, M. A., Orbell, S.J. and Mockett, A.P.A. 1995. Protection provided by a commercial available vaccine against different strains of turkey rhinotracheitis virus. *Veterinary Record*, **136**: 392-393.
- Cook, J. K. A. 1997. Respiratory diseases in chickens, including the significance of avian pneumovirus. Simposio Facta, Brazil.
- Cook, J, K. A., Huggins, M. B., Orbell, S. J., and Senne, D. A. 1999. Preliminary antigenic characterization of an avian pneumovirus isolated from commercial turkeys in Colorado, USA. *Avian Pathology*, **28**, 607-617.
- Cook, J. K.A. 2000. Avian rhinotracheitis. *Revue Scientifique Technic, OIE*, 19: 602-613.
- Gough, R.E. 2003. Avian pneumovirus. In: Diseases of poultry. 11<sup>th</sup> ed. Y. M. Saif, H. J. Barns, A. M. Fadly, J. R. Glisson, L. R. McDougald and D. E. Swayne. Eds. Iowa State University Press, Ames, Ia, USA. Pp 92-99.
- Hafez, H.M. and Lohren, U. 1990. Swollen head syndrome: clinical observations and serological examinations in West Germany. *Deutsche Tierarztliche Wochenschrift*, **97**: 322-324.
- Jones, R.C., Nayler, C.J., Bradbury, J.M., Savage, C.E., Worthington, K., and Williams, R.A. 1991. Isolation of turkey rhinitracheitis -like virus from broiler breeder chickens in England. *Veterinary Record*, **129**: 509-510.
- Morley, A. J. and Thomson, D.K. 1984. Swollen head syndrome in broiler chickens. *Avian Diseases*, **28**: 238-243.

- Owoade, A. A., Dukatez, M. F., and Muller, C.P. 2006. Seroprevalence of avian influenza virus, infectious bronchitis virus, reovirus, avian pneumovirus, infectious laryngotracheitis virus and avian leukosis virus in Nigerian poultry. *Avian Diseases*, **50**: 222-227.
- Pattison, M., Chettle, N., randall, C.J., and Wyeth, P.J. 1989. Observations on swollen head syndrome in broiler and broiler breeder chickens. *Veterinary Record*, **125**: 229-231.
- Paul, McMullin. 1998. Diagnosis, management and control of avian pneumovirus infection in broiler parent chickens. Facta Symposium, Campinas, Brazil.
- Seal, B. 2000. Avian penumovirus and emergence of a new type in the United States of America. *Animal Health and Research Review*, **1**: 67-72.